

Morphometric Study of a Hybrid Population of *Acacia verniciflua* and *A. aspera* (*Acacia* subgenus *Phyllodineae*)

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Abstract

Hybrids between the common variant of *Acacia verniciflua* A.Cunn. and *Acacia aspera* Lindl. are reported from central Victoria, in remnant vegetation in the median strip of the Hume Highway, south of Benalla. Plants of both species and suspected hybrids were sampled and mapped along a belt transect. Morphological pattern analysis, based largely on phyllode characters and using ordination and agglomerative classification techniques, confirmed a hybrid population of plants intermediate between *A. verniciflua* and *A. aspera* and possible backcrosses to one or other of the parent species. Scanning electron microscopy revealed that hybrids had density of simple hair and morphology of glandular trichomes intermediate between the two parents. The site has been disturbed, which may account for the hybrid swarm.

Introduction

Acacia verniciflua A.Cunn. (Varnish Wattle) occurs in South Australia, Queensland, New South Wales, Victoria, and Tasmania Fig. 1 (Fig. 1; Maslin 2001). It is a viscid shrub to small tree (1–8 m high) and highly variable across its range, with four variants in Victoria: common, southern, Casterton and Bacehus Marsh (Entwistle *et al.* 1996). Phyllodes are characterised by two main veins and glands (presumably resin-secreting), which are stalkless or sunken.

Acacia aspera Lindl. (Rough Wattle) is a spreading viscid shrub (0.5–2 m high) found in New South Wales and Victoria. It is less variable than *A. verniciflua* (Maslin 2001) with a single, newly recognised subspecies, *Acacia aspera* subsp. *parviceps*, found in

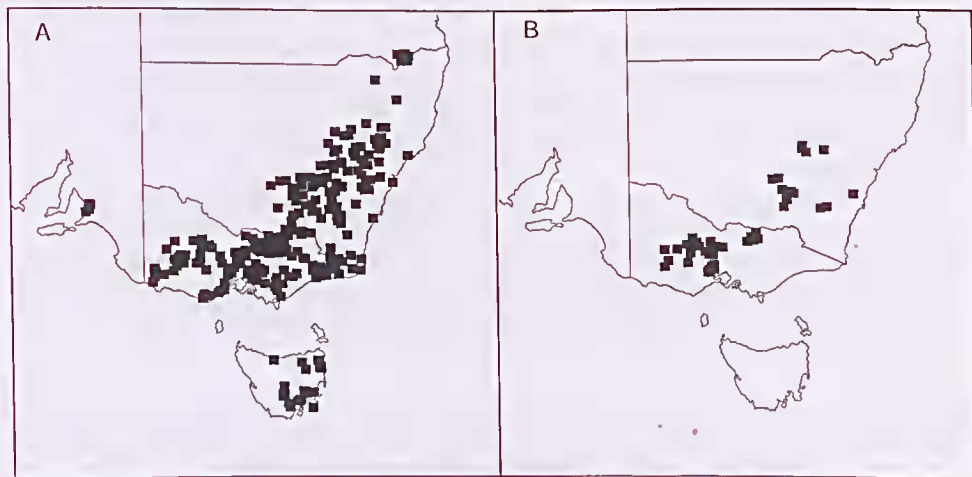


Figure 1. Geographic distribution of *Acacia verniciflua* and *A. aspera* in south-eastern mainland Australia.



Figure 2. Location of the study site, the Hume Highway, south of Benalla, Victoria.

Victoria in the Brisbane Ranges and near Beaufort (Walsh, 2004). *Phyllodes* have one main vein and flower buds have acuminate exserted bracteoles (Entwisle *et al.* 1996). The species is characterised by numerous simple hairs and glandular trichomes covering vegetative and reproductive structures. The latter are variously described as glandular hairs (Boughton 1989) or coarse hairs, gland-tipped (Entwisle *et al.* 1996).

Both *A. verniciflua* and *A. aspera* are classified in section *Phyllodineae* and are considered related (Maslin 2001). Entwisle *et al.* (1996) placed *A. verniciflua* in Group 3 and *A. aspera* in Group 4 of their treatment, although these groups are informal and do not necessarily reflect natural relationships. The geographic ranges of the two species overlap and in some localities they are sympatric. A mixed-species population of the common variant of *A. verniciflua*, *A. aspera*, and plants intermediate in morphology, has been found in central Victoria, south of Benalla (Fig. 2). The vegetation is remnant eucalypt woodland in the median strip of the Hume Highway and the site appears to have been disturbed (e.g. soil and drainage) by road-making. Given that *A. verniciflua* and *A. aspera* flower at the same time of the year, between July and November (Entwisle *et al.* 1996), it was hypothesised that the intermediate plants are of hybrid origin between these two putative parents.

Natural hybridisation in *Acacia* Miller has been reported by a number of authors (Cheel 1935; Moffett 1956; Leach & Whiffin 1978; Ali & Qaiser 1980; Sedgley *et al.* 1992; Entwisle *et al.* 1996). Hybrids between *A. aspera* and *A. montana* have been recorded from the Whipstick, near Bendigo, Victoria (Maslin 2001), but no record of hybridisation between *A. verniciflua* and *A. aspera* has been reported to date.

Method

The study site is located 185km north of Melbourne along the Hume Highway, 3 km north east of Erreys road, on the median strip between the north- and south-bound lanes (Latitude: S 36° 36' 11", Longitude: E 145° 49' 30"). The remnant vegetation is dominated by *Eucalyptus microcarpa* (Maiden) Maiden, *E. melliodora* A.Cunn. ex

Schauer, *E. polyanthemus* Schauer, *E. blakelyi* Maiden and *E. macrorhyncha* F.Muell. ex Benth. *Acacia* species at the site include *A. verniciflua*, *A. aspera*, *A. pycnantha* Benth. and *A. genistifolia* Link, and grasses are predominantly *Austrodanthonia* spp.

A 40 m belt transect line running north-south, parallel with the road, was used to sample the *A. aspera* and *A. verniciflua* population in the median strip. Sampling was done in April 2003, and no flowers or legumes were present. Some plants showed signs of severe drought stress. Every healthy plant within 2m to the east of the transect line was sampled. The height of the plant and the distance along the transect line where it was found were recorded. A sample of branchlets with phyllodes was taken from each plant for morphological measurements. Plants were sampled for voucher herbarium specimens (RP1-35 lodged at MELU). One plant typical of each of *A. aspera* and *A. verniciflua* and four suspected hybrids were also sampled for phyllode preservation in 70% ethanol.

Thirty-five specimens collected from the study site were measured for 12 phyllode, branchlet and bud characters (Table 1). Forty additional specimens from the National Herbarium of Victoria were also measured (Appendix 1, numbered 36 - 75), consisting of twenty specimens each of *Acacia verniciflua* and *A. aspera*, and including 10 samples from the north-east of Victoria. Five phyllodes were measured from each specimen, while bud and branchlet characters were scored by scanning the entire specimen. Averages of the measurements were taken in order to produce an individual score for each specimen.

Table 1. Characters measured for morphological analysis.

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1. Phyllode length (mm)
 2. Phyllode width at widest point (mm)
 3. Phyllode curvature (falcate or not): widest distance of inner edge of phyllode from a vertical line between base and tip of phyllode (mm)
 4. Distance from base of phyllode to the widest point of phyllode (mm)
 5. Distance from base of phyllode to the marginal gland (mm)
 6. Phyllode thickness (mm)
 7. Number of main veins on phyllode
 8. Length of pulvinus (mm)
 9. Resin glands stalked or sessile
 10. Branchlets distinctly tomentose (hairs) or not
 11. Flower buds with bracteoles exerted or not
 12. Peduncle and buds hairy or not
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The computer package PATN (Belbin 1995) was used for phenetic pattern analysis of the data. The Manhattan metric distance measure was used to construct a distance matrix and both unweighted pair group method of averaging (UPGMA) and non-metric multidimensional scaling (NMDS) were used for classification and ordination analyses (Clifford and Stephenson 1975).

Samples of the phyllodes preserved in 70% ethanol were prepared for Scanning Electron Microscopy (SEM). Samples were gradually dried out through a series of 70%, 80%, 90% and 100% ethanol. Sections were critical point dried, mounted onto stubs with an adhesive carbon disc, sputter coated with a thin coating of gold and viewed under a X1-30 Philips SEM.

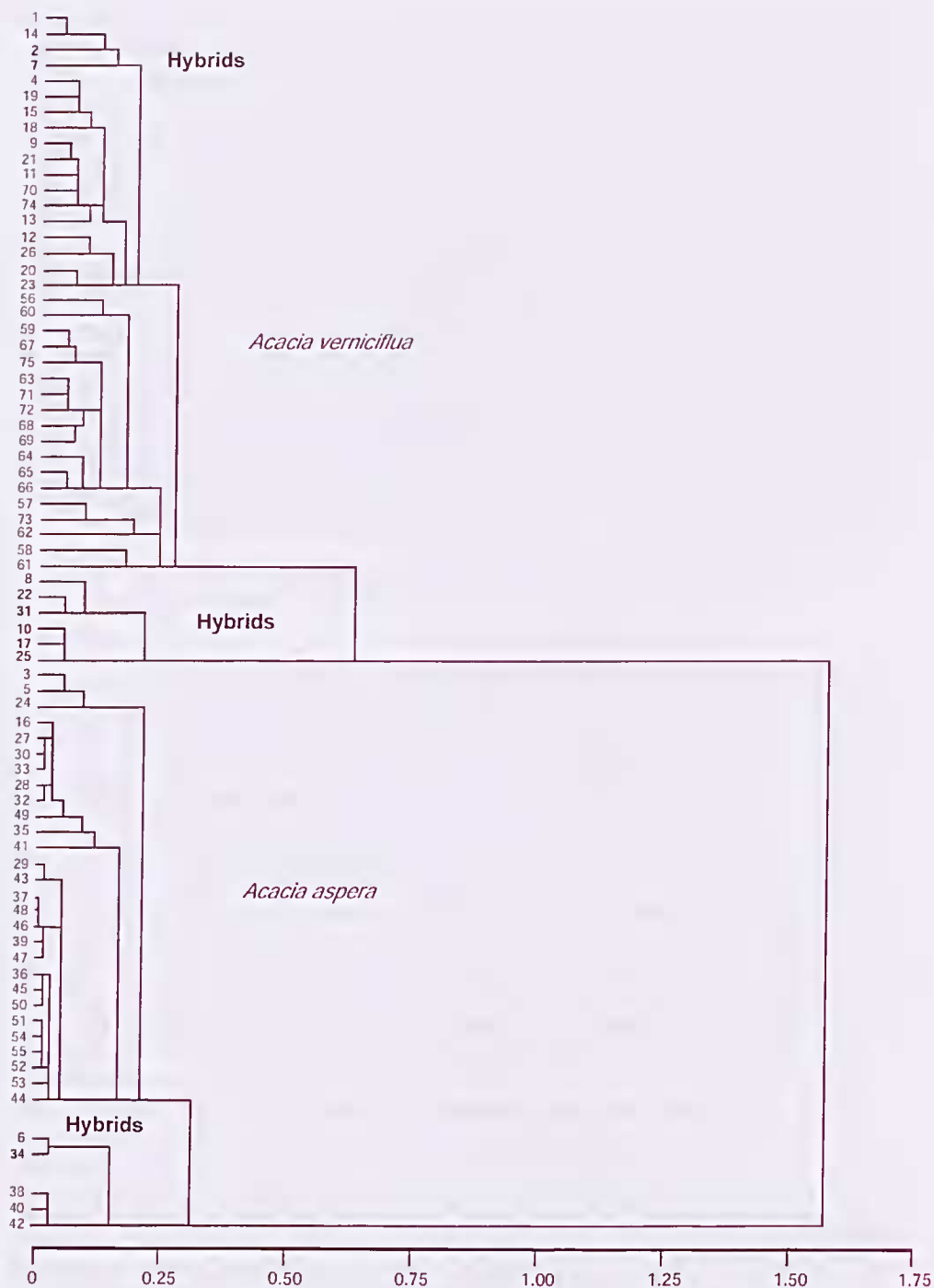


Figure 3. A dendrogram from the agglomerative cluster analysis of 75 specimens (1-35 from the filed site and 36-55 herbarium specimens at MEL). Suspected hybrids are shown in bold.

Results

The agglomerative classification (Fig. 3) of all 75 plants (field and herbarium samples) shows three main groups. The groups correspond to *A. verniciflua*, *A. aspera* and a cluster of six intermediate hybrid plants (specimens 8, 10, 17, 22, 25 and 31). This hybrid group clusters at a higher level in the dendrogram with *A. verniciflua* rather than with *A. aspera*. Specimens 2 and 7, categorised in the field as possible hybrids, cluster in the *A. verniciflua* group. Another two specimens thought to be hybrids, 6 and 34, cluster with a subgroup of *A. aspera* (herbarium specimens 38, 40 and 42) that have some phyllodes with two veins in addition to the more typical one-veined phyllodes.

The ordination scatter plot shown in three dimensions (Fig. 4) confirms the three main groups of the dendrogram. All ten suspected hybrids fall in between the two species groups of *A. verniciflua* and *A. aspera*, with specimens 2 and 7 being most like *A. verniciflua* and 6 and 34 being nearer to *A. aspera*. The three herbarium specimens of *A. aspera* from north-east Victoria that had some phyllodes with two veins and some with one vein, form a small cluster next to the main *A. aspera* group.

Phyllode length, width and measure of falcate shape display clearly the intermediacy

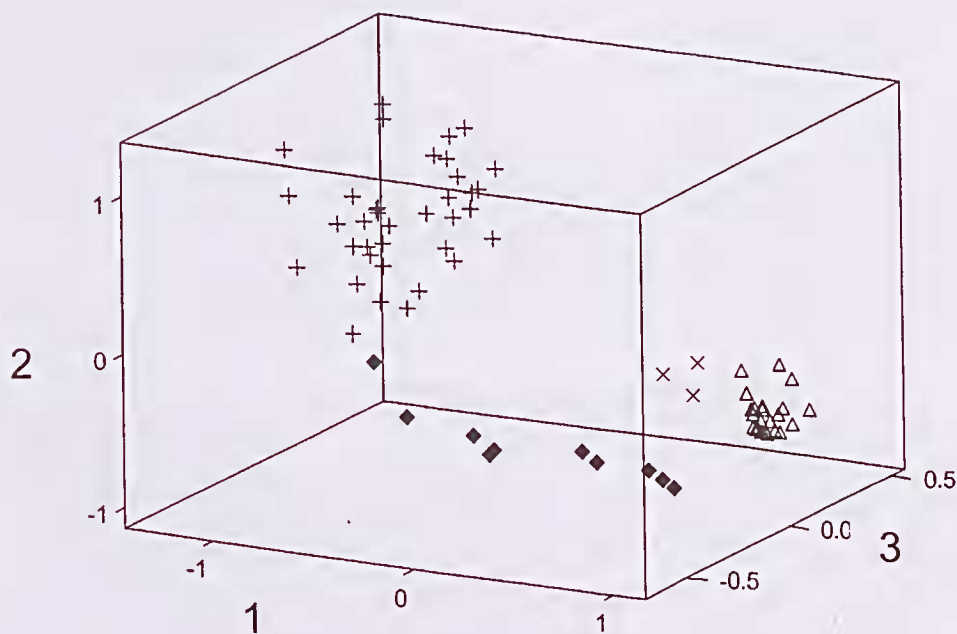


Figure 4. Three dimensional scatter-plot of the 75 specimens. +: *A. verniciflua* group, diamonds: hybrids, triangles: *A. aspera*, x: three herbarium specimens of *A. aspera* from north-east Victoria found to have phyllodes with both 1 and 2 veins.

of the hybrid specimens (Table 2, Fig. 5). The number of main veins for most of the hybrids was variable within the one plant, with some phyllodes having two veins and others only one vein. Whether bracteoles are exerted or not was variable between plants but constant within a plant. The intermediacy of the hybrids is clearly evident in the distribution of unicellular hairs and morphology of glands and glandular trichomes.

Typical of *A. aspera* (Fig. 6a, b) are multicellular glands raised well above the phyllode surface at the tip of a stalk. This contrasts with the presumably homologous but sunken glands of *A. verniciflua* (Fig. 6g), whilst the hybrids had either sessile or shortly stalked glands (Fig. 6c, d, e). The unicellular simple hairs of *A. aspera* are abundant on phyllodes, present but scattered sparsely on the hybrid phyllodes and absent from *A. verniciflua* (Fig. 6a, c, f).

Table 2. Diagnostic characters for parent species and hybrids. Mean values and range (minimum and maximum values) are given.

| Character | <i>A. aspera</i> | hybrids | <i>A. verniciflua</i> |
|---------------------------|------------------|----------------------------|-----------------------|
| Phyllode length (cm) | 1.8 (1.4–2.5) | 2.6 (2.3–3.1) | 4.8 (2.5–8.5) |
| Phyllode width (mm) | 3 (2–4) | 3 (3–3.5) | 5 (3–9) |
| Phyllode curvature (mm) | 0 straight | 1 intermediate | 2–3 subfalcate |
| Number of main veins | 1 | 1 & or 2 | 2 |
| Glands stalked or sessile | stalked | sessile or shortly stalked | sessile-sunken |
| Braeteoles exerted or not | + | +/- | - |

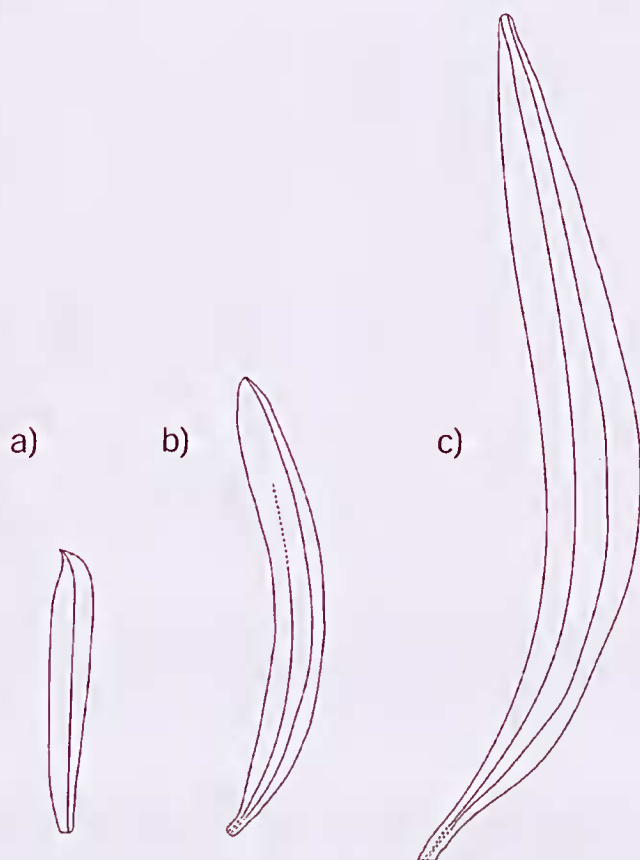


Figure 5. Phyllode shapes of (a) typical *A. aspera* (specimen 35), (b) an intermediate (putative F_1 hybrid, specimen 10) and (c) typical *A. verniciflua* (specimen 14).

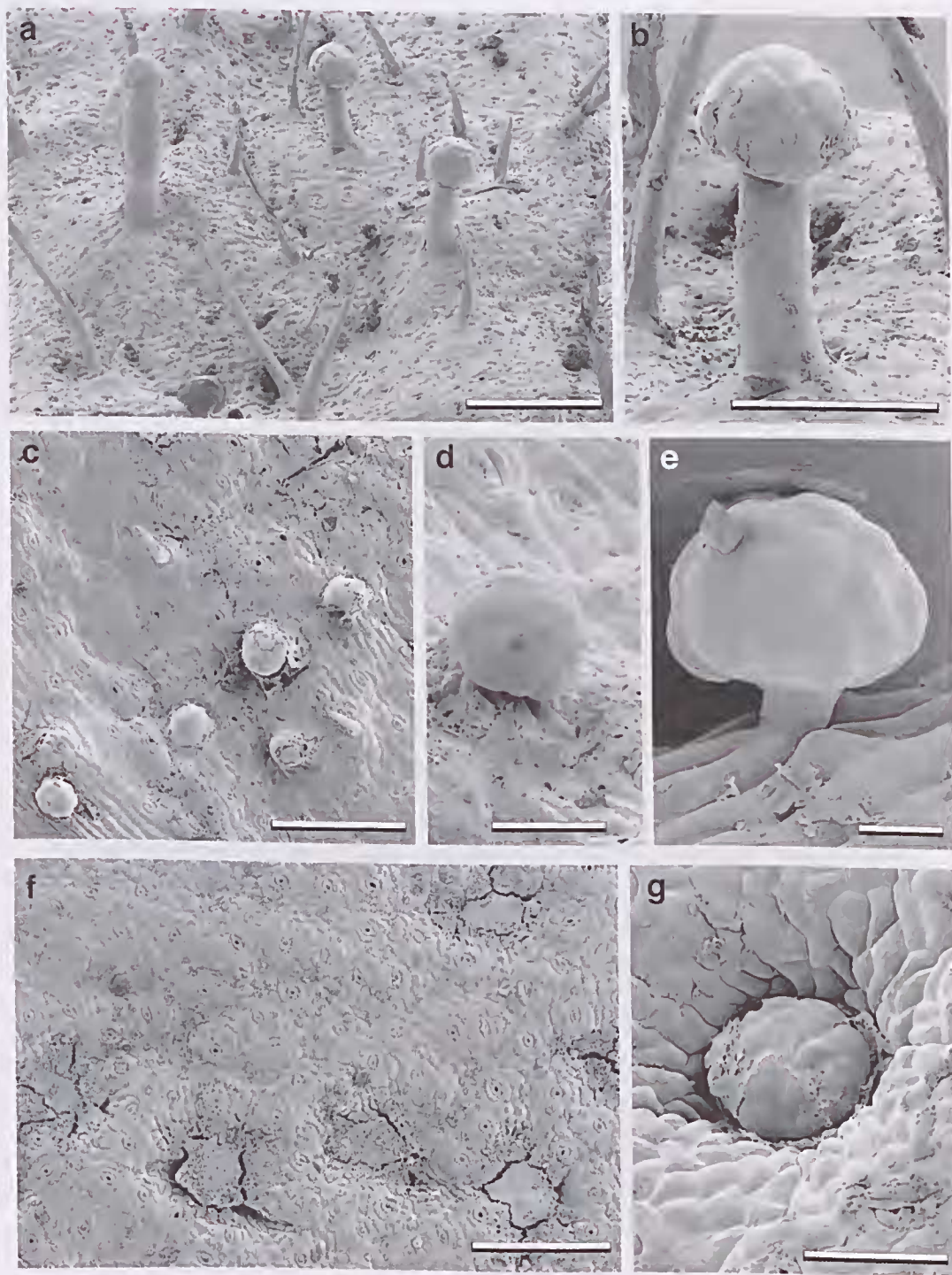


Figure 6. Scanning electron micrographs of the phyllode surfaces of *A. aspera* (specimen 35), a hybrid (specimen 31) and *A. verniciflua* (specimen 23). (a) *A. aspera* stalked resin glands and dense hairs, scale bar = 200mm, (b) at higher magnification, scale bar = 100mm; (c) Hybrid glands and some short hairs of hybrid, scale bar = 200mm, (d) and (e) at higher magnifications, with scale bars =50 and 20mm respectively; (f) *A. verniciflua* with sunken glands covered by resin, which has not dissolved, scale bar = 200mm, (g) sunken gland, scale bar = 50mm.

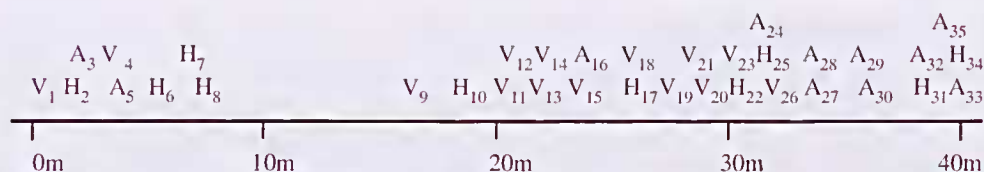


Figure 7. A diagram of the 40m belt transect line showing the relative position of plants sampled in this study; V = *A. verniciflua*, H = hybrids and A = *A. aspera*. Specimen numbers are given beside letters.

Figure 7 illustrates the distribution of the hybrids and the two parent species collected along the transect line at the study site. *Acacia aspera* and *A. verniciflua* are mixed over the study area with hybrids distributed evenly along the 40 metre transect line. For the most part Figure 7 shows that where the hybrids occur there are both *A. aspera* and *A. verniciflua* plants present within a short distance.

Discussion

The morphometric analysis has shown clearly the intermediacy of the suspected hybrid specimens. This supports the hypothesis that six of the field collected specimens are in fact hybrids of *A. aspera* and *A. verniciflua*, probably F_1 progeny. The two suspected hybrids (specimens 2 and 7) that were shown to be more similar to *A. verniciflua* than to the intermediate hybrids may be backcross F_2 progeny. The same explanation is offered for the two suspected hybrids (specimens 6 and 34) that were shown to be more similar to the *A. aspera* group. It is of interest that they were most similar to three herbarium specimens from north-east Victoria that had some phyllodes with two veins, which is suggestive of another instance of *A. aspera* \times *A. verniciflua* hybridisation. What controls the development of one or two veins in a single plant requires further examination.

Due to the timing of this study it was not possible to sample flowers, pods or seeds. It is therefore not possible to conclude from this study the fecundity of the hybrids. If the hypothesis that there has been a degree of back crossing is accepted, it would indicate that there is a degree of fertility amongst F_1 progeny.

A link between natural hybridisation and disturbance has long been established (Anderson 1948; Ali & Qaiser 1980; Sedgely *et al.* 1992; Mayer & Mesler 1993; Hawkins *et al.* 1999). Anderson (1948) put forward the theory of hybridisation of the habitat, which describes the need for an intermediate environment for the survival of the F_1 progeny, and a large amount of variation in the habitat for the survival of a variable F_2 generation. Disturbance of a habitat through the construction of roads may create the micro sites that allow for the establishment of a hybrid swarm (Mayer & Mesler 1993). Such disturbance can create less shading, more water and alter the soil of the area, creating an array of different conditions in which hybrids may grow. The site of the *Acacia* hybrids in the median strip of the Hume Highway is a disturbed site, sunken below the level of the road, acting as a drainage channel. A second site of a possible hybrid swarm further north-east along the Hume highway near Glenrowan has yet to be confirmed.

Conclusion

This study has demonstrated the hybrid status of intermediate plants in a mixed population of *A. verniciflua* and *A. aspera*. This hybrid is a new record for Victoria. Both *A. verniciflua* and *A. aspera* are widely distributed, so the 'genetic purity' of the species is not under great threat. Unless the area is cleared the hybrid population should persist at the site.

Acknowledgments

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Appendix 1

| <i>Acacia</i> species | Specimen No. | Collection No. | MEL No. | Location |
|-----------------------|--------------|----------------------------|------------|--|
| <i>aspera</i> | 36 | Beaglehole, A.C. 93423 | MEL2022434 | 10km S.S.E. of Winton |
| <i>aspera</i> | 37 | Beaglehole, A.C. 66150 | MEL618560 | Reef Hill Park |
| <i>aspera</i> | 38 | Canning, E.M. 1153 | MEL500736 | Between Benalla and Wangaratta on Hume Hwy |
| <i>aspera</i> | 39 | Hall, N. 79/98 | MEL605683 | Between Benalla and Wangaratta on Hume Hwy |
| <i>aspera</i> | 40 | Hall, N. 79/98 | MEL1516433 | Hume Highway near Glenrowan |
| <i>aspera</i> | 41 | Beaglehole, A.C. 81493 | MEL679129 | Warby Ranges |
| <i>aspera</i> | 42 | Bartolome, M. 193 | MEL2155846 | 16km E.S.E. of Benalla |
| <i>aspera</i> | 43 | Fletcher, R.J. 281 | MEL2022280 | Reef Hill Park |
| <i>aspera</i> | 44 | Piesse, A.D.J. 328 | MEL1551519 | 13km N.E. of Benalla on Hume Hwy |
| <i>aspera</i> | 45 | Rossiter, N.T. 1 | MEL616041 | N.E. of Glenrowan |
| <i>aspera</i> | 46 | Wallace, G. s.n. | MEL666606 | Tarnagulla |
| <i>aspera</i> | 47 | Beaglehole, A.C. 69018 | MEL618561 | 4km S.S.E. of Castlemaine |
| <i>aspera</i> | 48 | Muir, T.B. 1246 | MEL1500071 | 15 Miles N.E. of Stawell |
| <i>aspera</i> | 49 | LeBreton, C. s.n. | MEL234340 | Lerderderg Gorge |
| <i>aspera</i> | 50 | Clarke, I.C. 2027 | MEL1586231 | 3km W.S.W. of Elphinstone |
| <i>aspera</i> | 51 | Walter, C. s.n. | MEL500720 | Bendigo |
| <i>aspera</i> | 52 | Beaglehole, A.C. 50184 | MEL520336 | 8.5km W.S.W. of Inglewood |
| <i>aspera</i> | 53 | Beaglehole, A.C. 50254 | MEL520337 | 27 km N.E. of Stawell |
| <i>aspera</i> | 54 | Beaglehole, A.C. 38930 | MEL500735 | Lightning Hill Reservoir, Bendigo |
| <i>aspera</i> | 55 | Beaglehole, A.C. 18928 | MEL500731 | Llanelly |
| <i>verniciflua</i> | 56 | Court, A.B. & Willis, J.H. | MEL1529029 | 5 miles E. of Wilby |
| <i>verniciflua</i> | 57 | Fletcher, R.J. 263 | MEL2021041 | 30km S.E. from Wangaratta |
| <i>verniciflua</i> | 58 | Piesse, A.D.J. 426 | MEL1555746 | 10.4km S. of Whitfield |
| <i>verniciflua</i> | 59 | Willis, J.H. s.n. | MEL502697 | Central Summit of Mt. Ida, near Heathcote |
| <i>verniciflua</i> | 60 | Crawford, I. 1816 | MEL712592 | Dookie Agriculture College |
| <i>verniciflua</i> | 61 | Fletcher, R.J. 287 | MEL2022274 | Chiltern State Park, near Cyanide Dam |
| <i>verniciflua</i> | 62 | Fletcher, R.J. 288 | MEL2022273 | Chiltern State Park, near Cyanide Dam |
| <i>verniciflua</i> | 63 | Beaglehole, A.C. 80343 | MEL679039 | Boweya Flora & Fauna Reserve |
| <i>verniciflua</i> | 64 | Fletcher, R.J. 289 | MEL2022272 | Chiltern State Park, near Cyanide Dam |
| <i>verniciflua</i> | 65 | Beaglehole, A.C. 92378 | MEL2022424 | Snowfields, Mt Buffalo Reference Area |
| <i>verniciflua</i> | 66 | Beaglehole, A.C. 16962 | MEL1516197 | Mt. Victory, Grampians |
| <i>verniciflua</i> | 67 | Beaglehole, A.C. 16964 | MEL563430 | Mt. Victory, Grampians |
| <i>verniciflua</i> | 68 | Parfett, S.T.W. 56 | MEL1537955 | 200m S.S.E. of Scout Camp, Grampians |
| <i>verniciflua</i> | 69 | Beaglehole, A.C. 17456 | MEL563429 | Grampians |
| <i>verniciflua</i> | 70 | Unknown | MEL1529069 | Cultivated |
| <i>verniciflua</i> | 71 | Hicks, A.J. s.n. | MEL1529027 | Rodes Gap, Grampians |
| <i>verniciflua</i> | 72 | Muir, T.B. 2146 | MEL1529049 | 32 miles S. of Horsham |
| <i>verniciflua</i> | 73 | Court, A.B. s.n. | MEL1516049 | Near Chiltern |
| <i>verniciflua</i> | 74 | Walter, C. s.n. | MEL1528783 | Dandenong Ranges |
| <i>verniciflua</i> | 75 | Walter, C. s.n. | MEL1528778 | Dandenong Ranges |